

11P.11 Using EPR up close and from afar: Elucidating mechanisms in haem copper oxidases

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Haem copper oxidases constitute the terminal complex of the respiratory chain and catalyse the reduction of oxygen to water. This exergonic redox reaction is coupled to proton pumping across the inner mitochondrial or bacterial membrane. O₂ reduction occurs at the binuclear haem-Cu_B centre. Despite high resolution X-ray crystallographic structures, the properties of the catalytic redox states of the metal centres and their relation to protonation states within this class of enzyme remain still poorly understood. Modern EPR techniques (also in combination with magneto-optical studies) enable us to probe different catalytic intermediate states either directly or indirectly. From afar pulsed ELDOR spectroscopy, a technique for accurately measuring inter spin distances in the range 2–8 nm, is used to resolve subtle structural changes when applied to spin-labelled systems trapped in different intermediate states (e.g. P, R & F states) and which allows the study of local conformational changes in great detail. Using this technique conformational change within the proton uptake channels is discussed. Up close both EPR and magneto-optical techniques (Magnetic Circular Dichroism) are used to address the nature of the metal ligands in the binuclear centre as well as transiently formed radical species from different intermediate states as well as in oxidases from different species.

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11P.12 Fourier transform infrared spectroscopy reveals water molecules reorganization in cytochrome c oxidases

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The overall mechanism of electron transfer and the oxygen reduction chemistry in cytochrome c oxidases (CcO) are fairly well understood. However, the associated proton transfer pathways and the way protons are gated either to the binuclear center or to a site for translocation remains unclear. One feasible mechanism involves a network of water molecules that reorganize to protonically connect a conserved glutamic acid (E242 in bovine CcO) either to the binuclear center or to a trap site above the hemes on the proton translocation pathway (Wikström M *et al.*, 2003, *Biochim. Biophys. Acta* **1604**: 61–65). Water molecules have been resolved by X-ray crystallography both in the D channel that leads to E242 and also in the region above the hemes through which translocated protons might be expected to pass. However, none of these changed markedly between oxidized and reduced forms and water molecules that could connect E242 and the binuclear center or the proton trap site have never been observed. Such water networks are H-bonded chains that can have both strongly and weakly H-bonded –OH groups. The weakly H-bonded groups absorb in the infrared spectrum between 3500 and 3800 cm^{–1}. We used FTIR difference spectroscopy to detect such weakly H-bonded –OH groups that might change organization during catalysis in bovine CcO.

Complex spectral changes between 3680 and 3560 cm^{–1} were observed on diatomic ligand binding to the reduced binuclear center, a reaction that mimics the catalytic oxygen binding step. Their sensitivities to D₂O and H₂¹⁸O media confirmed that they arose from water molecules. Redox difference spectra also exhibited simpler changes in this region at 3674, 3619 and 3607 cm^{–1}. These transitions can be correlated to changes in the environment of a protonated carboxyl group that has been assigned to E242. Similar patterns of water reorganization can be observed in other CcO homologues suggesting that they are caused by a common conserved mechanism. The data are discussed in relation to possible functional roles in proton/electron coupling.

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11P.13 Photo-dependent binding structures of CO and NO on the heme-copper site in bovine cytochrome c oxidase

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Cytochrome c oxidase (CcO) catalyzes the O₂ reduction on the heme a₃-Cu_B site. The binding of O₂ on the ferrous heme a₃ iron (Fe_{a3}) as the sixth ligand results in formation of the catalytic intermediate as O₂-bound form. The O₂ analogues, carbon monoxide (CO), nitric oxide (NO) are also bound as the sixth ligand. The cuprous Cu_B is coordinated by three histidine imidazoles and the fourth coordination position is empty. Previous infrared studies reported that CO photo-dissociated from Fe_{a3} is transiently bound to Cu_B before the rebinding to Fe_{a3} in a temperature dependent manner [1, 2]. As in the case of CO, NO can be photo-dissociated from Fe_{a3} and rebound to Fe_{a3}, though the rebinding rate of NO is faster than that of CO [3, 4]. Here, we report photo-dependent binding structures of CO and NO in bovine CcO analyzed by absorption spectra and X-ray structures under low temperatures [5, 6]. The observed absorption spectral changes of the CcO crystals indicate that CO and NO are irreversibly photodissociated from Fe_{a3} at the temperatures of 100 K and 50 K, respectively. X-ray structures determined at above temperatures under light illumination revealed that both CO and NO were similarly bound at the fourth coordination position of Cu_B by the side-on manner. The CO- and NO-bound Fe_{a3} structures have been also determined at the temperatures of 280 K and 100 K, respectively. These results suggest that the photo-dissociated forms of CO and NO are stabilized nearby Cu_B under low temperatures. To directly demonstrate this explanation, we determined the NO-binding geometry at 50 K in the dark. The X-ray structure showed that NO remained bound to Fe_{a3} in the dark.

References

- [1] J.O. Alben, P.P. Moh, F.G. Fiamingo, R.A. Altschuld, *Proc. Natl. Acad. Sci. U. S. A.* **78** (1981) 234–237.
- [2] Ó. Einarsson, R.B. Dyer, D.D. Lemon, P.M. Killough, S.M. Hubig, S.J. Atherton, J.J. Lopez-Garriga, G. Palmer, W.H. Woodruff, *Biochemistry* **32** (1993) 12013–12024.
- [3] S. Yoshida, H. Hori, Y. Oori, *J. Biochem.* **88** (1980) 1623–1627.
- [4] R. LoBrutto, Y.-H. Wei, S. Yoshida, H.L. Van Camp, C.P. Scholes, T.E. King, *Biophys. J.* **45** (1984) 473–479.
- [5] K. Ohta, K. Muramoto, K. Shinzawa-Itoh, E. Yamashita, S. Yoshikawa, T. Tsukihara, *Acta Cryst. F66* (2010) 251–253.